Analytical Evidence Report

HPLC-UV results for the submitted GHK-Cu sample

REPORT NUMBER
THPN-GHK-100-092126

GENERATED
October 21, 2026

GHK-Cu

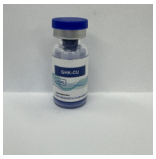
Sample vial 4, injection 1 | Client report THPN-GHK-100-092126

Results and chromatograms

Lot # NXP-GHKCU-02 | Client: NexPept

Laboratory Analysis: LAB-CA-02

Laboratory identity and applicable credentials are maintained by Testides and are available to regulatory authorities where required by law.

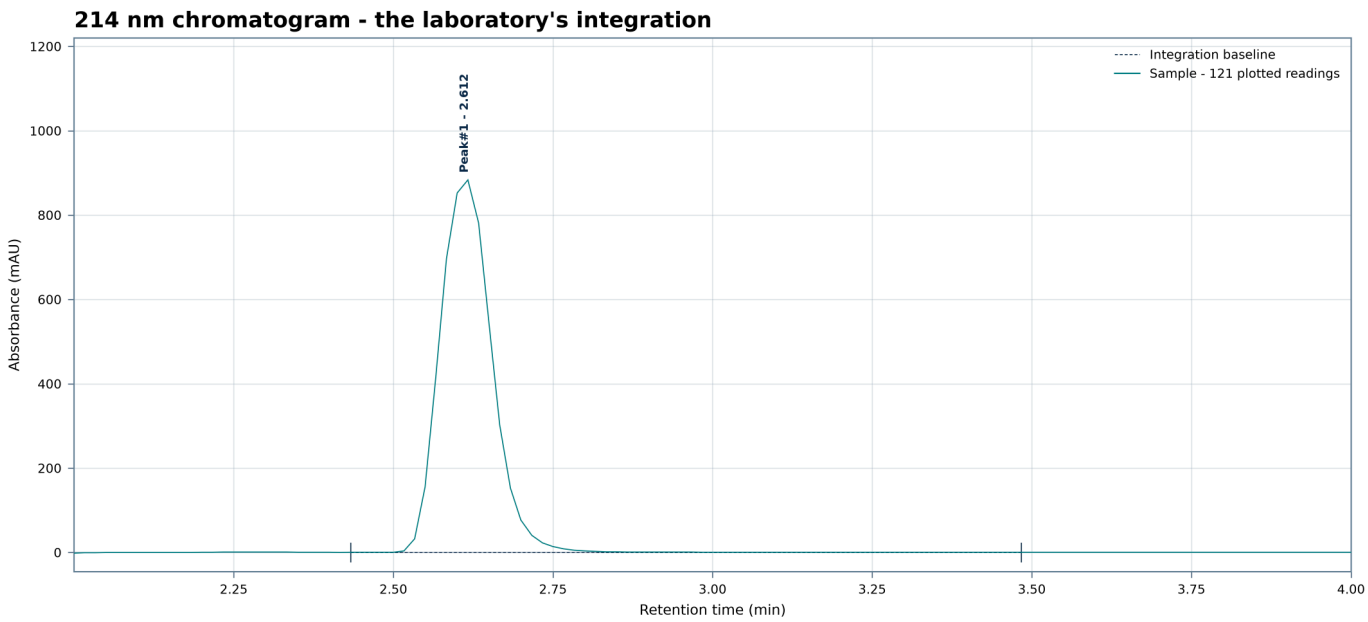
VIAL PHOTOGRAPH	SAMPLE INFORMATION		ANALYSIS		RUN DETAILS	
	Peptide:	**GHK-Cu**	Method:	**HPLC-UV**	Run date:	**Sep 25, 2026**
	Client ID:	**THPN-GHK-100-092126**	Channel:	**214 nm**	Acquired:	**16:27:26 UTC**
	Label claim:	**100 mg**	Integration:	**Laboratory**	Run time:	**15 min**
	Injection:	**10 uL, 214 nm**	Identity:	**Conforms**	Instrument:	**HPLC 02**

RESULTS AT A GLANCE

PROJECTED AMOUNT	HPLC PURITY	CONTENT VS. LABEL	IDENTITY
131.44 mg†	**99.96%**	**131.4%**	**Conforms**
BASED ON SAMPLE PREPARATION	214 NM AREA PERCENT*	131.44 MG OF 100 MG CLAIMED	PEAK IDENTITY CONFIRMED**

CHROMATOGRAM

As integrated by the laboratory - every plotted reading, with its baselines and peak marks



HPLC PURITY (214 NM)

Main peak area %	Integrated peaks	Smallest peak integrated	Recovered chromatogram
99.96%	**1**	**100.00%**	**±0.01%**
Laboratory integration	Counted in the total	Of total area	Area vs. laboratory table

Purity is the main peak's share of the total peak area the laboratory integrated at 214 nm. The laboratory integrated no other peak; it does not state a reporting threshold, so a peak smaller than it integrates is not counted. Integrating the chromatogram recovered from the laboratory's report reproduces its main-peak area to within 0.01%. This result rests on a single injection.

The chromatogram is recovered point for point from the laboratory's report: 121 plotted readings from 2.00 to 4.00 min, joined as drawn; nothing is smoothed or resampled.

* HPLC purity is the main peak's share of all peak area the laboratory integrated; every peak is itemised on page 3.

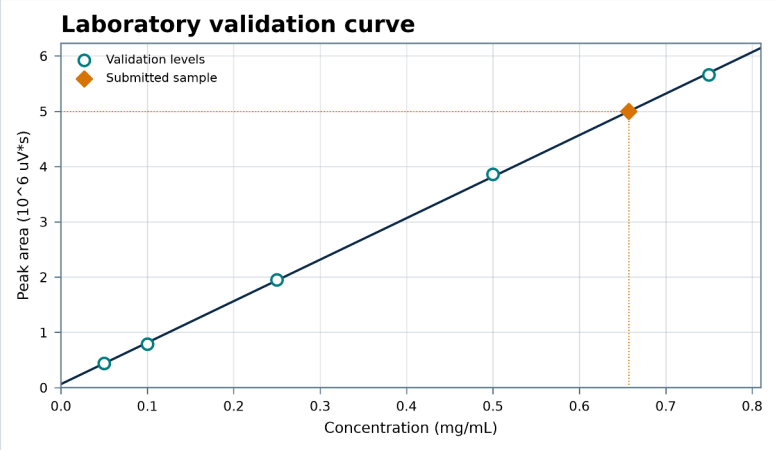
** Identity conforms. The reference standard ran on a different method from the sample, so retention times are not compared.

GHK-Cu

Sample vial 4, injection 1 | Client report THPN-GHK-100-092126

Reference comparison and calculation
Lot # NXP-GHKCU-02 | Client: NexPept

CONCENTRATION CALIBRATION - 214 NM (LABORATORY VALIDATION CURVE)

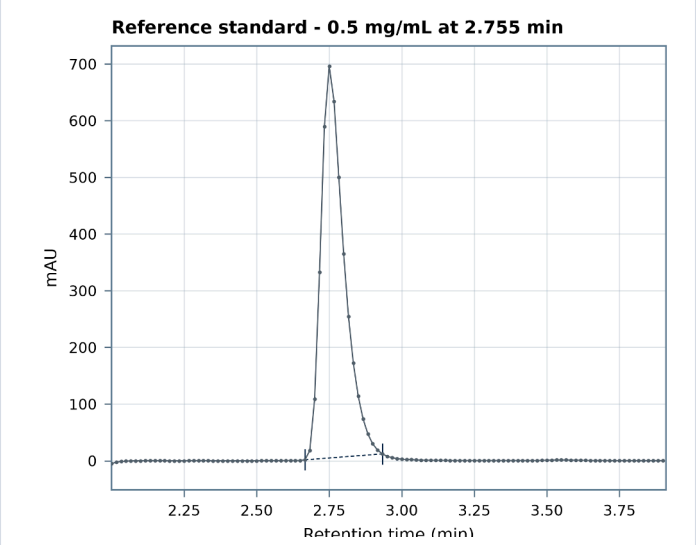


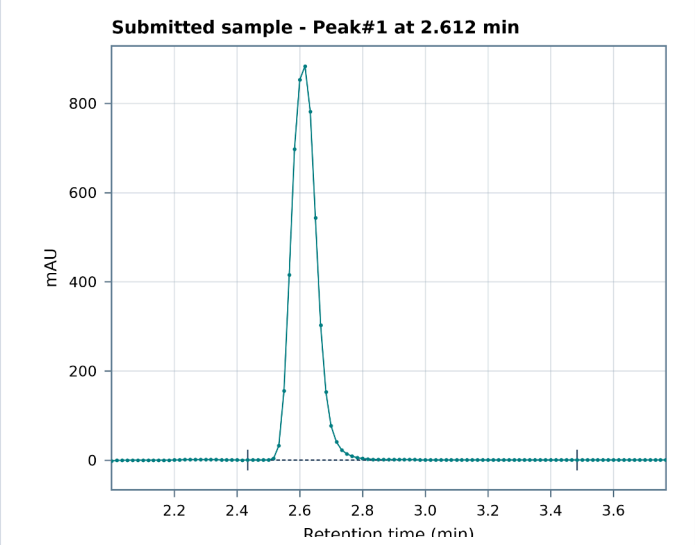
mg/mL	Area (uV*s)	Recovery
0.75	5,662,587	99.44%
0.5	3,863,237	101.23%
0.25	1,953,897	100.72%
0.1	787,791	96.48%
0.05	437,225	99.58%

Area = 7,507,610 C +63,436
Unweighted OLS through 5 levels | R2 0.99979
Sample area 4,997,591 = 0.6572 mg/mL (label: 0.5)
x 2 mL x 100 = 131.44 mg

† 2 mL diluent, diluted x100 to the working 0.5 mg/mL
Validated 2026-08-21 | HPLC 02 | injector RSD 0.43% (n=6) | reference 99.2% pure

MAIN PEAK COMPARED WITH THE LABORATORY'S REFERENCE STANDARD





Native plotted readings with the laboratory's integration baselines and peak marks; nothing is smoothed or resampled.
Reference standard injected 2026-08-18 during the laboratory's validation of this curve on another method than the sample's (LC02_Peptide15min_AD_Jul15_26), so retention times differ.

BACTERIAL ENDOTOXINS (USP <85>)

RESULT	THRESHOLD	METHOD	CONTROLS
PASS	**≤ 1 EU/mL**	**Gel-clot LAL**	**Passed**
NEGATIVE BY THE LABORATORY	AT OR BELOW PASSES	USP <85>	POSITIVE AND NEGATIVE

At or below the 1 EU/mL threshold

The measured value, the gel-clot readings, the lysate, the controls and the calculation are on page 3.

GHK-Cu

Sample vial 4, injection 1 | Client report THPN-GHK-100-092126

Definitions, calculation detail, and scope

Lot # NXP-GHKCU-02 | Client: NexPept

HOW TO READ THIS REPORT

Start with the result on page 1. Pages 2 and 3 show the curve, comparison, and peaks behind it.

01 Recovered chromatogram
The laboratory's trace, read back point for point from its report.

02 Integrated peak table
Every peak the laboratory integrated, and which one is the analyte.

03 Validation curve
Converts the peak area to a concentration for this analyte.

WHAT THE RECOVERED CHROMATOGRAM ADDS

How the chromatogram on this report was obtained

How it was recovered

The laboratory's data system draws the chromatogram in its report as vector data. Every vertex it drew was read back - 121 readings - and both axes were calibrated from the report's own tick marks. Integrating the recovered trace over the laboratory's baselines and peak marks reproduces the area in its peak table to within 0.01%.

What it can and cannot show

It shows every peak, shoulder and baseline the laboratory integrated, at the resolution the laboratory printed. It carries a single wavelength and no UV spectrum, so it cannot reveal a co-eluting compound the way diode-array data can.

Scope: the recovered chromatogram reproduces the laboratory's report. It does not re-measure the sample.

HOW THE PURITY RESULT WAS MEASURED

214 nm area percent, from the laboratory's own integration

The laboratory integrated each peak to its own baseline and named the analyte's peak in its processing method. Purity is that peak's share of the total integrated area; the shares below are recomputed from the areas, and each peak's area is checked against the recovered chromatogram.

Purity (%) = assigned peak area / total integrated peak area x 100

Peak	RT (min)	Area (uV*s)	Area %	Tailing	Plates	Role	Recovered
1	2.612	4,997,591	99.96	1.17	4,926	Peak#1 - main	4,997,965 (+0.01%)

Integrated peaks 1 | smallest 100.00% of total | injections 1

Area percent assumes every peak absorbs at 214 nm like the main peak; the analyte's peak is the one the laboratory's method names. Recovered areas are compared for the analyte's peaks of 1% or more.

BACTERIAL ENDOTOXINS — HOW THE RESULT WAS MEASURED

Measured: 0.5 EU/mL | laboratory result: Negative

Replicate	1/2	1/4	1/8	1/16	1/32	Endpoint
1	+	+	-	-	-	1/4
2	+	+	-	-	-	1/4

Each replicate at each two-fold dilution of the sample solution: + firm gel, - no gel

Lysate PYROGENT 125 Plus lot DC0423; labelled sensitivity λ = 0.125 EU/mL. Dilutions 1/2 to 1/32, two-fold, in 2 replicates.

Endotoxin concentration = lysate sensitivity / mean endpoint = 0.125 EU/mL / 1/4 = 0.5 EU/mL. The mean endpoint is the geometric mean of the replicates' endpoints (the most dilute tube that gels).

Laboratory worksheet MC-02777, tested Sep 23, 2026, approved Sep 25, 2026. The positive control gelled and the negative control did not, as required.

WHAT THIS REPORT CAN AND CANNOT TELL YOU

The report explains the measurement; it does not decide how a material should be used

Observed evidence
The laboratory's chromatogram, its integration, and its validation curve.

Calculated results
Area-percent purity, concentration, projected amount, and calibration trace.

Method context
Retention time against the reference standard or the report's other vials, where on the same method.

Safety and regulatory status
No conclusion on safety, efficacy, fitness for use, or regulatory status.

This analytical record does not authorize or endorse manufacture, sale, marketing, distribution, or use of the material.